

Natural Products Synthesis

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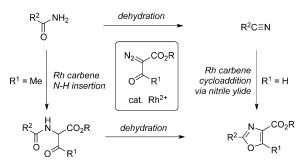
Total Synthesis of the Posttranslationally Modified Polyazole Peptide Antibiotic Plantazolicin A

Hiroki Wada, Huw E. L. Williams, and Christopher J. Moody*

Abstract: The power of rhodium-carbene methodology in chemistry is demonstrated by the synthesis of a structurally complex polyazole antibiotic. Plantazolicin A, a novel soilbacterium metabolite, comprises a linear array of 10 fivemembered rings in two pentacyclic regions that derive from ribosomal peptide synthesis followed by extensive posttranslational modification. The compound possesses potent antimicrobial activity, and is selectively active against the anthraxcausing organism. A conceptually different synthesis of plantazolicin A is reported in which the key steps are the use of rhodium(II)-catalyzed reactions of diazocarbonyl compounds to generate up to six of the seven oxazole rings of the antibiotic. NMR spectroscopic studies and molecular modeling reveal a likely dynamic hairpin conformation with a hinge region around the two isoleucine residues. The compound has modest activity against methicillin-resistant Staphylococcus aureus (MRSA).

n the one and a half centuries since August Kekulé and Archibald Scott Couper independently proposed that a carbon atom could form four bonds to other atoms (including other carbon atoms), [1,2] the existence of divalent carbon species with a six-electron valence shell has intrigued chemists. Once regarded as mechanistic curiosities or fleeting intermediates, such divalent species, now known as carbenes, have moved center stage as a result of the isolation of the first stable carbene in the late 1980s,[3] to be followed by the now familiar stable N-heterocyclic carbenes, [4] which as ligands have revolutionized transition-metal catalysis.^[5] Also metallocarbene intermediates, derived from diazo compounds, participate in a plethora of reactions that are useful in chemical synthesis.^[6] We now demonstrate the power of carbene chemistry in the synthesis of the structurally unique, complex polyazole antibiotic plantazolicin A, in which up to six of the seven five-membered oxazole rings of the natural product are formed from simple precursors, such as carboxamides or nitriles, facilitated by carbene methodology (Scheme 1).[7-10]

Plantazolicin A (1) and plantazolicin B (2) are novel metabolites isolated from the soil bacterium *Bacillus amyloliquefaciens* FZB42.^[11,12] The structures consist of a linear



Scheme 1. Synthesis of oxazoles from carboxamides and nitriles via rhodium carbenes.

array of five-membered rings (azoles) that derive biosynthetically from amino acids by ribosomal peptide synthesis followed by wide-ranging posttranslational modification. [13-15] However, it is the potent antimicrobial activity of plantazolicin A against Gram-positive organisms that has attracted much attention. In particular, the compound is selectively active against the anthrax-causing organism Bacillus anthracis (Sterne).[11,13] Although the polyazole nature of plantazolicin A is highly reminiscent of the thiopeptide antibiotics, [16-18] a synthetic derivative of which has entered the clinic against Clostridium difficile infections, [19,20] there are key differences. Specifically, the linear nature of the antibiotic with its two pentacyclic regions represents a challenge for chemical synthesis that, in combination with the antimicrobial activity, makes plantazolicin A highly attractive for further study. The first total synthesis of plantazolicin A was reported by Süssmuth and co-workers in 2013, [21] and very recently, a second total synthesis was reported by Ley and co-workers.^[22] Both syntheses relied on classical peptide coupling. We now report a conceptually different synthesis of plantazolicin A on the basis of carbene chemistry.

Our strategy was to construct the azole rings by using carbene intermediates, and hence our retrosynthetic analysis divided the molecule into two fragments, 3 and 4, each adorned with appropriate protecting groups, with the sensitive oxazoline ring in the C-terminus right-hand fragment 4 to be formed by a late-stage cyclodehydration reaction (Scheme 2). In contrast with other approaches, we elected to introduce the guanidine moiety later in the synthesis. Thus, the starting point for our synthesis was the known ornithinederived thiazole-4-ester 5, [23] which was readily converted into the corresponding carboxamide 6 to set the scene for our first step involving a carbene (Scheme 3). The key metallocarbene N-H insertion was carried out by heating a mixture of methyl 2-diazo-3-oxobutanoate and amide 6 in the presence of rhodium(II) acetate dimer (2.5 mol%) in dichloromethane in a microwave reactor (200 W, 80 °C), to give the ketoamide

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Scheme 2. Retrosynthetic analysis of plantazolicin A.

7, which was immediately cyclized to oxazole 8 by cyclodehydration with triphenylphosphine and iodine in the presence of a base. [24] The difficulties associated with the isolation of the product from triphenylphosphine oxide were largely overcome by the use of polymer-supported triphenylphosphine. The ester 8 was converted into the corresponding carboxamide 9 and then into the thiocarboxamide 10 by treatment with the Lawesson reagent. A Hantzsch reaction formed the next ring and delivered the three-azole array 11 (Scheme 3). With two further oxazole rings to install, we instigated an iterative carbene carboxamide N-H insertioncyclodehydration sequence. Thus, ester 11 was converted into carboxamide 12, which underwent the desired carbene insertion to give ketoamide 13, the cyclodehydration of which gave tetra-azole 14 in 52% yield over two steps. A second iteration via carboxamide 15 and ketoamide 16 produced the desired penta-azole 17, although the yield of this last ring-forming step was poor.

In view of the unsatisfactory formation of the fifth and final ring in the penta-azole 17, we sought an alternative, while still satisfying our desire to use carbene methodology to construct the azole rings. Thus, the protected threoninamide 18 underwent rhodium(II)-catalyzed N-H insertion followed by cyclodehydration to give oxazole ester 20 in good yield, the deprotection of which resulted in oxazole 21 with a threoninederived free amine (Scheme 4). This amine was then coupled with the thiazolecarboxylate formed by simple hydrolysis of the bis(thiazolyl)oxazole 11, thus resulting in the formation of the linear tetra-azole 22. Subsequent ring closure of the threonine fragment by the use of DAST methodology gave oxazoline 23,^[25] the dehydrogenative aromatization of which with bromotrichloromethane in the presence of a base^[26] yielded the previously prepared penta-azole 17.

With an improved route available to the key penta-azole, the synthesis rapidly progressed towards the complete lefthand fragment 3 of the antibiotic. Acid-mediated removal of the Boc group allowed for installation of the dimethylamino group by a reductive amination reaction with formaldehyde to give 25, which underwent cleavage of the remaining Nprotecting group in preparation for the introduction of the 2trimethylsilylethoxycarbonyl (Teoc)-protected guanidine

moiety onto amine 26. To this end, we developed a new reagent, the pyrazole carboxamidine 27, which successfully incorporated the protected guanidine to give penta-azole 28. Hydrolysis of the ester finally gave the desired fragment 3 (Scheme 4) in 1.2% overall yield over 17 steps.

The presence of four C5-unsubstituted oxazoles in the Cterminal fragment of the antibiotic, in contrast to the three 5methyloxazoles present in the N-terminal fragment 3, necessitated a slight change in methodology. Thus, the order of steps was reversed, and the carboxamide was initially dehydrated to the nitrile, with subsequent direct conversion into the oxazole in the rhodium-catalyzed carbene cycloaddition step (Scheme 1).[10,27] Hence, the synthesis of fragment 4 started with the known isoleucine-isoleucine dipeptide 29, [28] which was converted into the carboxamide 30 and then the required nitrile 31 (Scheme 5). These transformations set the stage for the direct installation of the first oxazole 32 by a dirhodium(II)-catalyzed reaction with the formyl diazo compound ethyl 2-diazo-3-oxopropanoate, although a change in catalyst to dirhodium tetrakis(heptafluorobutyramide) was beneficial. [27] The sequence was iterated to deliver the bisoxazole ester 35 by the conversion of oxazole ester 32 into nitrile 34 and thereafter a second rhodium-carbene step. Meanwhile, nitrile 37 also underwent rhodium-catalyzed carbene cycloaddition to give oxazole 38,[10] which was deprotected to give amine 39 in preparation for union with the bisoxazole fragment. Bisoxazole ester 35 was hydrolyzed to acid 36, and subsequently coupled to amine 39 by use of the HBTU protocol to give adduct 40. A DAST-mediated cyclodehydration and aromatization completed the linear array 41 of four oxazole rings, and this ester was subsequently hydrolyzed to the corresponding carboxylic acid 42 for further elaboration (Scheme 5). Separately, Boc-Phe-OH was protected as its 2-trimethylsilylethyl (TMSE) ester 43, the Boc group was cleaved, and the ensuing amine 44 was united with N-Boc-protected allothreonine to give dipeptide 45. The dipeptide was deprotected in acid, and the resulting amine 46 underwent coupling with the tetra-oxazole acid 42 to give, after removal of the Boc group, the complete Cterminal fragment 4 in 1.7% overall yield over 13 steps from the isoleucine dipeptide 29.

15363



Scheme 3. Synthesis of intermediate penta-azole 17: a) 35% aqueous NH₃, EtOH, room temperature, 93%; b) methyl 2-diazo-3-oxobutanoate (1.5 equiv), rhodium(II) acetate dimer (2.5 mol%), CH₂Cl₂, 80°C, microwave (200 W); c) polymer–Ph₃P (1.6 equiv), I₂ (1.6 equiv), Et₃N (3.2 equiv), CH₂Cl₂, room temperature, 72% (2 steps); d) 35% aqueous ammonia, MeOH, THF, room temperature, 80%; e) Lawesson reagent (0.7 equiv), CHCl₃, room temperature, 55%; f) ethyl bromopyruvate (5.0 equiv), KHCO₃ (10 equiv), DME, -10° C; g) trifluoroacetic anhydride (5 equiv), 2,6-lutidine (10 equiv), DME, -10° C; h) K₂CO₃ (5.0 equiv), EtOH, H₂O, room temperature, 71% (3 steps); i) 35% aqueous NH₃, EtOH, room temperature, 84%; j) methyl 2-diazo-3-oxobutanoate (1.4 equiv), rhodium(II) acetate dimer (2.5 mol%), CHCl₃, 60°C; k) polymer–Ph₃P (3.5 equiv), I₂ (3.5 equiv), Et₃N (7.0 equiv), CH₂Cl₂, room temperature, 52% (2 steps); l) LiOH, MeOH/THF/H₂O (1:5:5), room temperature, 93%; m) EtO₂CCl, Et₃N, THF, 35% aqueous NH₃, room temperature, 57%; n) methyl 2-diazo-3-oxobutanoate (1.5 equiv), rhodium(II) acetate dimer (2.5 mol%), CHCl₃, 60°C; o) polymer–Ph₃P (4.0 equiv), I₂ (4.0 equiv), Et₃N (8.0 equiv), CH₂Cl₂, room temperature, 11% (2 steps). Boc = tert-butoxycarbonyl, Cbz = benzyloxycarbonyl, DME = 1,2-dimethoxyethane.



Scheme 5. Synthesis of right-hand tetra-azole fragment 4: a) 35% aqueous ammonia, MeOH, THF, room temperature, 76%; b) DBU (5.0 equiv), ethyl dichlorophosphate (3.0 equiv), CH₂Cl₂, 0°C, 80%; c) ethyl 2-diazo-3-oxopropanoate (3.0 equiv), rhodium(II) perfluorobutyramide dimer (2.5 mol%), CHCl₃, 60°C, 53%; d) 35% aqueous ammonia, EtOH, THF, room temperature, 89%; e) DBU (5.0 equiv), ethyl dichlorophosphate (3.0 equiv), CH₂Cl₂, 0°C, 79%; f) ethyl 2-diazo-3-oxopropanoate (3.0 equiv), rhodium(II) perfluorobutyramide dimer (2.5 mol%), CHCl₃, 60°C, 59%; g) LiOH (58 equiv), EtOH/THF/H₂O (1:5:5), room temperature, 65%; h) ethyl 2-diazo-3-oxopropanoate (3.0 equiv), rhodium(II) perfluorobutyramide dimer (2.5 mol%), CHCl₃, 60°C, 51%; i) 2 m HCl in ether, room temperature, quantitative; j) HBTU (1.5 equiv), 39 (1.5 equiv), Et₃N (2.0 equiv), CH₂Cl₂/DMF (1:1), room temperature, 76%; k) DAST (1.7 equiv), K₂CO₃ (5.0 equiv), CH₂Cl₂, -78°C, quantitative; l) BrCCl₃ (4.0 equiv), DBU (4.0 equiv), CH₂Cl₂, 0°C, 59% (2 steps); m) LiOH (29 equiv), EtOH/THF/H₂O (1:5:5), room temperature, 76%; n) 4 m HCl in dioxane, room temperature, 56%; o) Boc-protected allothreonine (1.0 equiv), HBTU (2.0 equiv), 44 (2.0 equiv), Et₃N (4.0 equiv), CH₂Cl₂/DMF (1:1), room temperature, 65%; p) 4 m HCl in 1,4-dioxane, room temperature, 79%; q) HBTU (1.5 equiv), 46 (1.5 equiv), Et₃N (3.0 equiv), CH₂Cl₂/DMF (1:1), room temperature, 58%; r) 4 m HCl in 1,4-dioxane, room temperature, quantitative; s) HBTU (1.8 equiv), 3, Et₃N (3.7 equiv), CH₂Cl₂/DMF (1:1), room temperature, 50%; t) DAST (30.0 equiv), CH₂Cl₂, -78°C, 52%; u) TASF (30.0 equiv), DMSO, room temperature; v) HFIP, room temperature, 31%. DMF = N,N-dimethylformamide, DMSO = dimethyl sulfoxide, TMSE = 2-trimethylsilylethyl, TASF = tris (dimethylamino) sulfonium difluorotrimethylsilicate, HFIP = hexafluoroisopropanol.

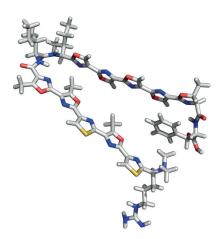
Having successfully obtained both the N- and C-terminal halves of the antibiotic, we coupled polyazoles **3** and **4** in the presence of HBTU and Et₃N to give **48**. The threonine moiety of the intermediate **48** was cyclized in the presence of DAST to afford the oxazoline **49**, the ¹H NMR and ¹³C NMR

spectroscopic data of which matched those of an intermediate prepared in an earlier synthesis. [21] All that remained was to remove the three silicon-based protecting groups, and in common with previous workers, we found that the removal of these groups was not completely straightforward, and



required a two-stage strategy involving the sequential use of two fluoride-based reagents (Scheme 5). Following purification by HPLC, the NMR spectroscopic data of our synthetic material matched those reported for the natural antibiotic (see Tables S1 and S2 in the Supporting Information); the compound also co-eluted on HPLC with authentic material (see Figures S1–S3). Our synthetic material also exhibited modest potency against methicillin-resistant $\it Staphylococcus aureus$ (MRSA) with a minimum inhibitory concentration (MIC) of > 33 $\mu g m L^{-1}$ (compare with the value of > 128 $\mu g m L^{-1}$ found for the natural material $^{[13]}$).

Finally, we were interested in the conformation of plantazolicin A. NMR spectroscopy NOESY and TOCSY experiments were carried out (see the Supporting Information) along with molecular modeling. The lack of long-range NOEs suggests a moderately dynamic molecule with rigid oxazole/thiazole arms that are not in close contact for any appreciable time. However, the strong NOEs around the central two isoleucine residues suggest that this portion of the molecule could act as a hinge region leading to a dynamic hairpin conformation. A structure consistent with these data is shown in Figure 1.



 $\label{eq:Figure 1.} \textbf{Most probable conformation of plantazolicin A on the basis of NMR spectroscopy and molecular modeling.}$

The carbene-based synthesis described above is not only a practical route that could be used to provide further quantities of the fascinating antimicrobial agent plantazolicin A, but it also makes available a wide range of fragment structures and analogues that are not made available by Nature's biosynthetic machinery. The full biological evaluation of these novel structures is under way.

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